Agrisera
Part of Olink® Group

This product is **for research use only** (not for diagnostic or therapeutic use) **contact: support@agrisera.com**

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com

Product no AS22 4860

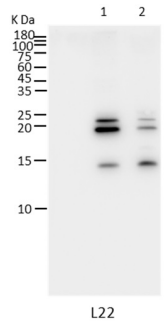
Anti-L22 | 50S ribosomal protein L22 (chloroplastic)

Product information

Immunogen	KLH-conjugated peptide derived from *Arabidopsis thaliana* L22 protein sequence. UniProt: P56795 TAIR: ATCG00810 and *Nicotiana tabacum* P06389
Host	Rabbit
Clonality	Polyclonal
Purity	Antigen affinity purified serum, in PBS pH 7.4
Format	Lyophilized
Quantity	50 µg
Reconstitution	For reconstitution, add 50 µl of sterile or deionized water.
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please, remember to spin tubes briefly prior to opening them to avoid any losses that might occur from lyophilized material adhering to the cap or sides of the tubes.

Application information

Recommended dilution	1 : 1000 - 1: 5000 (WB)
Expected | apparent MW	18.5 kDa
Confirmed reactivity	*Arabidopsis thaliana*
Predicted reactivity	Dicots, *Brassica napus*, *Hordeum vulgare, Nicotiana tabacum, Oryza sativa Solanum lycopresicum, Solanum tuberosum, Pisum sativum, Zea mays* Species of your interest not listed? Contact us
Not reactive in	*Chlamydomonas reinhardii*
Selected references	To be added when available, antibody available in February 2025.



Samples:

1. 20 µg of *Arabidopsis thaliana* seedlings extract of wildtype.
2. 20 µg of *Arabidopsis thaliana* seedlings extract of a mutant with a protein import defect

20 µg/well of total protein extracted freshly from 08-day-old Arabidopsis thaliana seedlings using 1% SDS and 8M urea. Proteins eluted in 8M urea (w/o SDS) were denatured with 2x Laemmli buffer (LB) at 75 °C for 7 min. Samples were separated using SDS-PAGE (20% gel) and blotted for 1.5 h to nitrocellulose (pore size of 0.45 µm), using wet transfer in the cold. Blot was blocked with 5% milk for ON/4ºC with agitation. Blot was incubated in the primary antibody at a dilution of 1:1000 for 7 h/4°C with agitation (5% Blotto in TBS-T). The antibody solution was decanted, and the blot was rinsed briefly twice and washed thrice for 15 min each in TBS-T at RT. Blot was incubated in anti-rabbit IgG HRP-conjugated secondary antibody diluted to 1:15 000 for 1 h/RT with agitation. The blot was washed thrice for 15 min each in TBS-T at RT and developed with the following chemiluminescent detection reagent: AS16 ECL-N-10 AgriseraBright. The exposure time was 30 seconds.

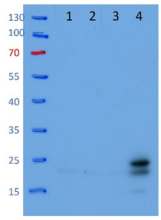
The middle band is L22 protein. Band pattern can vary depending upon analyzed material and applied protocol.

Courtesy of Project Associate. Kavyashree A and Dr. Naresh Loudya, Indian Institute of Science, Bengaluru, India

Agrisera
Part of Olink® Group

This product is **for research use only** (not for diagnostic or therapeutic use) **contact: support@agrisera.com**

Agrisera AB | Box 57 | SE-91121 Vännäs | Sweden | +46 (0)935 33 000 | www.agrisera.com



Samples:
1 - 15 ug of *Arabidopsis thaliana* whole seedling extract (Control)
2 - 15 ug of *Arabidopsis thaliana* whole seedling treated with drug-1 for 16hr
3 - 15 ug of *Arabidopsis thaliana* whole seedling treated with drug-2 for 16hr
4 - 15 ug of *Arabidopsis thaliana* whole seedling and enrich to chloroplast

15 µg/well of total protein extracted freshly from *Arabidopsis thaliana* 8-day-old seedlings. Exact buffer components were: 8M Urea buffer components at 4 °C. Samples were separated in the RT on 10 % SDS-PAGE and blotted for 2 h to PVDF (pore size of 0.2 µm), using: we transfer in the cold. Blot was blocked with 10 % milk for 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1: 3000 for 1h/RT with agitation in PBS with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in PBS at RT with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1: 5000 in for 1 h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent. Exposure time was 1 minute.

L22 protein band is a middle band, clearly visible in enriched chloroplast fraction. Longer exposure time will visualize bands in all samples.

Courtesy of Dr. Kuo-En Chen Department of Biology, Washington State University in St. Louis, USA